



Histological effects of toxoplasmosis and its treatments on male and female rats

Zahraa Sadoon Al-Ghezy¹, Fadhil Abbas Al-Abady¹, Ali Esmail Al-Snafi^{2*}

1. Department of Biology, College of Science, University of Thi qar - Iraq,

2. Department of Pharmacology, College of Medicine, University of Thi qar – Iraq.

ABSTRACT

The testis section of toxoplasma infected male rats showed vacular degeneration of spermatogonia and spermatocytes. The seminiferous tubules revealed sever degeneration, shrinkage, necrosis, hemorrhages with disappearance of epithelial lining of the majority of seminiferous tubules. They were also appeared separated with an irregular outlines and surrounded by fibrin. Seminiferous tubule lumen contained little amount of sperm with appearance of giant cells, polymorph nuclear leukocyte and exfoliated cellular debris. The epididymal sections of *Toxoplasma gondii* infected male rats treated by DMSO showed infiltration of mononuclear and multinucleated giant cell. Epididymal sections also showed hyperplasia of duct lining and contained toxoplasma cyst. Ovary sections of infected female rats treated by DMSO revealed morphological differences compared to the non infected group treated with DMSO, they showed decreased primary and secondary follicle with increased follicle atresia. Uterine section of infected female rats treated by DMSO showed hypertrophy of the endometrium and myometrium, polymorphic inflammatory infiltration, fewer glands, increased endometrial thickness with congestion of vessels. Pyrimethamine and sulphadiazine nor cause histological changed neither potentiate the histological effects of toxoplasmosis in infected and non infected male and female rats.

Keywords: *Toxoplasma gondii*, infection, male, female, rats, histology, pyrimethamine, sulphadiazine

*Corresponding Author Email: aboahmad61@yahoo.com

Received 23 March 2016, Accepted 28 March 2016

Please cite this article as: Ali AS *et al.*, Histological effects of toxoplasmosis and its treatments on male and female rats. American Journal of Pharmacy & Health Research 2016.

INTRODUCTION

Many studies correlated between reproductive performance and Toxoplasmosis¹⁻³. *Toxoplasma gondii* deteriorated the male and female reproductive performance in many experimental animals⁴⁻⁹. Toxoplasmosis decreased hypothalamic, pituitary and gonadal secretion^{1,3,8}. Furthermore, toxoplasmosis also caused profound adverse effect on human reproductive functions¹⁰. Acute toxoplasmosis infection caused hypogonadotrophic gonadal insufficiency in male patients regardless of the course of the disease¹¹. The women with toxoplasmosis may complain spontaneous abortions, stillbirths, intrauterine growth retardation, preterm deliveries, or fetal anomalies¹²⁻¹³.

T. gondii infection is of particular public health interest due to the ability of this parasite to cause infertility, therefore this study was designed to investigate the effect of toxoplasmosis on histology of male and female rats reproductive systems.

MATERIAL AND METHOD

The study was carried out on 42 males and 42 females rats (*Rattus norvegicus*) ranging in weight from 250 to 300g, all rats were housed in an air-conditioned animal room at an ambient temperature of 23 ± 2 C and in a 12h light / 12h dark cycle. Half of the males and females were infected intraperitoneally with 1×10^7 tachyzoites of *T. gondii* intraperitoneally¹⁴. Infected groups were examined for documentation of the infection with the using of real-time PCR. Infected groups (21 males and 21 females) and non infected group (21 males and 21 females) were divided into 3 subgroups (7 each) and treated with dimethyl sulphoxide (DMSO), sulphadiazine 12.5 mg/kg or pyrimethamine 200 mg/kg. Sulphadiazine and pyrimethamine were given in DMSO as a single oral daily dose for 60 days in males and 2 estrus cycles in females. At the end of the treatment period, testis, epididymis, of males and ovary and uterus of females were fixed in formalin for histological examination¹⁵.

RESULTS AND DISCUSSION

Testicular and epididymal sections:

The sections of the testis of non infected male rats treated with DMSO showed normal histological structures. The seminiferous tubules were intact with normal appearance of basement membrane, primary and secondary spermatocytes. The lumen was full with sperms, and with intact Sertoli cells. The seminiferous tubules appeared adjacent to each other with regular outlines. The interstitial tissues were intact. Also there were no histological changes in the non infected male rats treated with sulphadiazine or pyrimethamine.

The testis section of toxoplasma infected male rats showed vacular degeneration of spermatogonia and spermatocytes. The seminiferous tubules revealed sever degeneration, shrinkage, necrosis, hemorrhages with disappearance of epithelial lining of the majority of seminiferous tubules. They were also appeared separated with an irregular outlines and surrounded by fibrin. Seminiferous tubule lumen contained little amount of sperm with appearance of giant cells, polymorph nuclear leukocyte and exfoliated cellular debris. Sloughing of Leydig cells was also recorded.

The sections of the testis of infected male rats treated with sulphadiazine or pyrimethamine showed improvement of testicular structure. The section appeared with less or absent of vacular degeneration, necrosis and hemorrhages. The seminiferous tubules appeared with intact epithelial lining and the lumen full with sperms. The interstitial tissues also appeared intact with almost normal structure. It appeared that pyrimethamine was more effective than sulphadiazine in attenuation of the testicular histological changes associated with toxoplasmosis.

The epididymal sections of non infected male rats treated with DMSO, sulphadiazine, or pyrimethamine, showed that the epididymal duct was lined with an epithelium composed of principal and basal cells. The luminal diameter and the thickness of the peritubular smooth muscle increases from the proximal to the distal regions Few sperm were found in the initial segment, but a large mass of sperm aggregates were located in the cauda.

The epididymal sections of *Toxoplasma gondii* infected male rats treated by DMSO showed infiltration of mononuclear and multinucleated giant cell. Epididymal sections also showed hyperplasia of duct lining and contained toxoplasma cyst. Sperms appeared impacted in the lumen along with desquamated cellular debris. However, some epididymal ducts appeared atrophied with low amount of sperms.

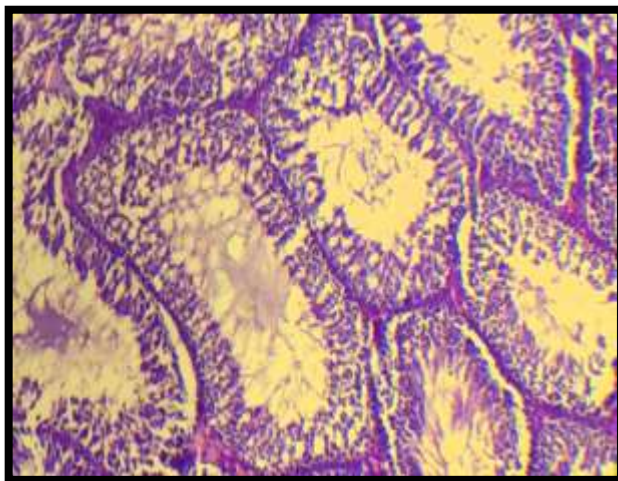
The epididymal sections of *Toxoplasma gondii* infected male rats treated with sulphadiazine, or pyrimethamine revealed hyperplasia of duct lining and slight degeneration. In the majority of the sections, the ducts appeared with normal diameter and epithelial lining, and the cavity was full with sperms.

Ovarian and uterine sections:

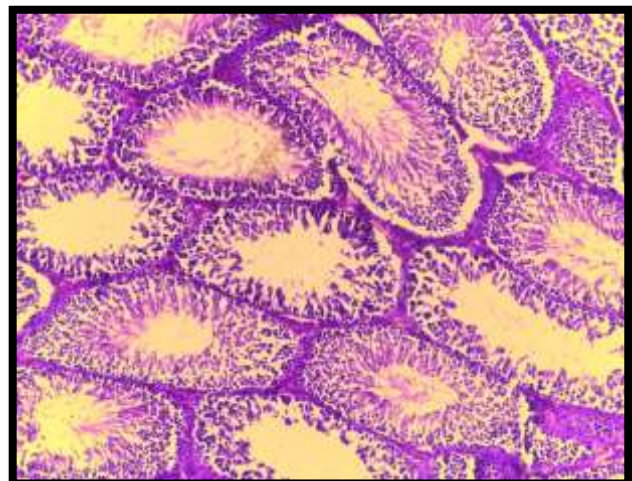
Ovary sections in non infected groups treated with DMSO, sulphadiazine, or pyrimethamine, showed normal histological pictures, the primary and secondary follicles and corpus luteum showed normal appearance. Ovary sections of infected female rats treated by DMSO revealed morphological differences compared to the non infected group treated with DMSO, they showed decreased primary and secondary follicle with increased follicle atresia, the atretic follicles were

characterized by degenerating oocytes, disorganized granulosa cell layers and folded zona pellucid. However, some of the primary follicles showed normal appearance. In the infected groups treated with sulphadiazine or pyrimethamine, the numbers of healthy primary and secondary follicles were increased with decreasing of the number of atretic follicles.

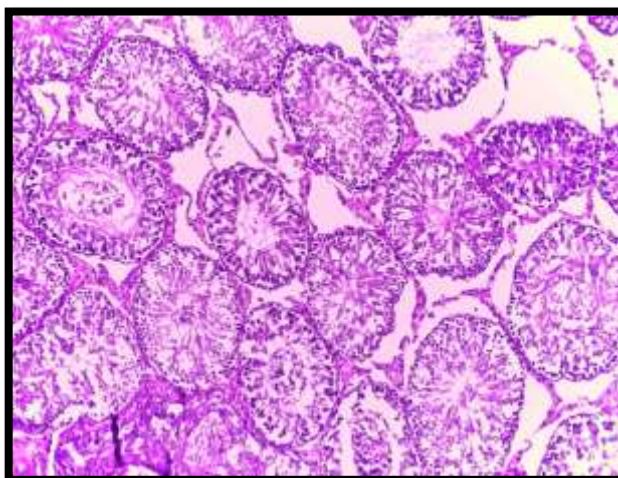
Uterine sections in non infected groups treated with DMSO, sulphadiazine, or pyrimethamine, showed normal histological pictures. The luminal and glandular epithelium were normal in appearance, normal thickness of endometrium and myometrium with low numbers of polymorphonuclear cells appeared within the lamina propria. Uterine section of infected female rats treated by DMSO showed hypertrophy of the endometrium and myometrium, polymorphic inflammatory infiltration, fewer glands, increased endometrial thickness with congestion of vessels in endometrium and myometrium. Uterine sections in infected groups treated with sulphadiazine, or pyrimethamine, also showed hypertrophy and increased endometrium and myometrium.



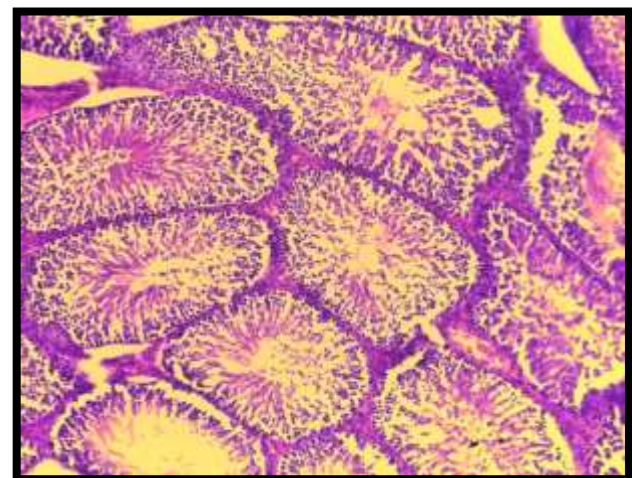
A



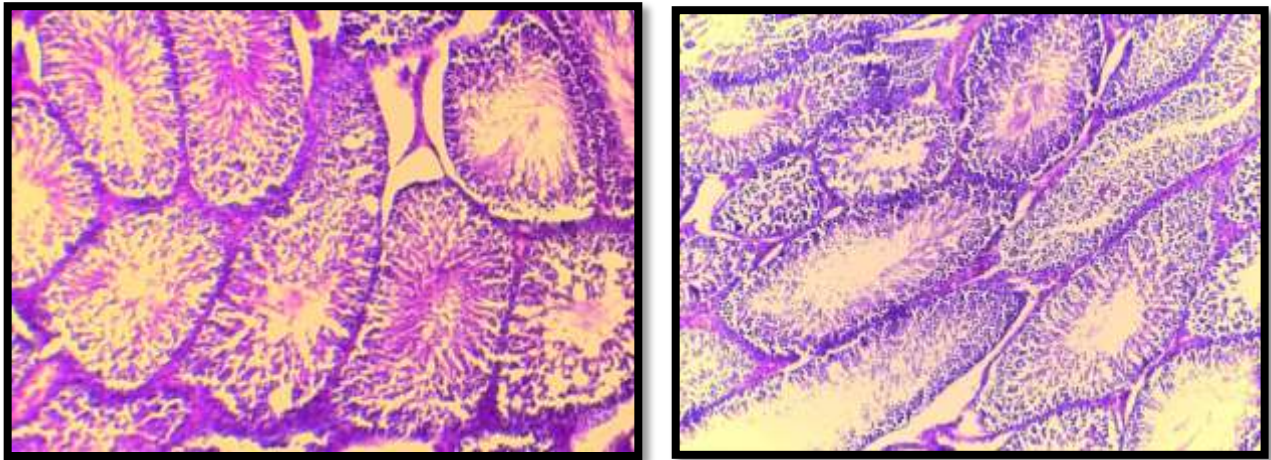
B



C



D

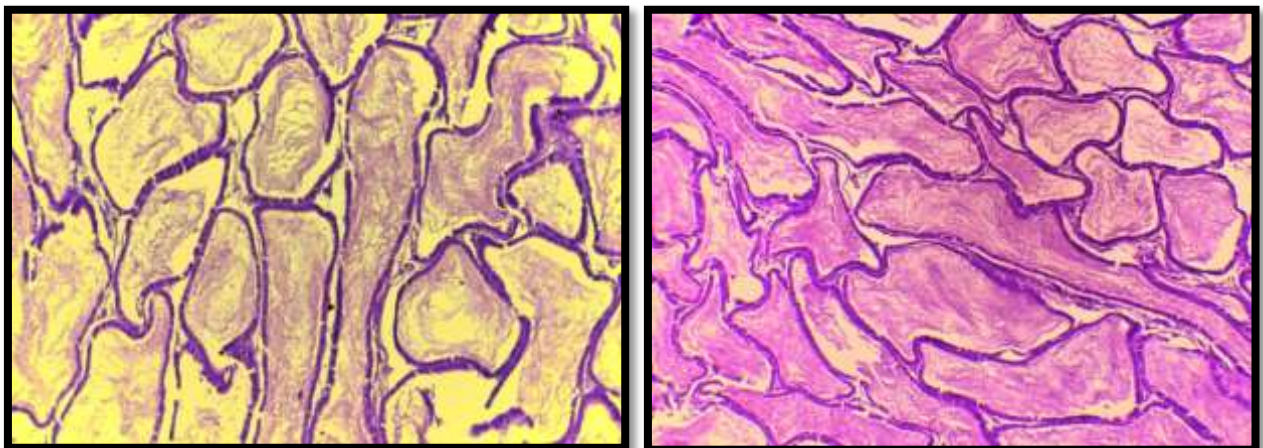


E

F

Figure 1: testicular section of non infected male rats treated with DMSO

(A), pyrimethamine (B) and sulphadiazine (C) showed the normal histological structure. The testicular section of infected male rats treated with DMSO showed tubular degeneration, shrinkage, disappearance of epithelial lining of seminiferous tubules. They were also appeared separated and surrounded by fibrin. with appearance of giant cells, polymorph nuclear leukocyte and exfoliated cellular debris (D). The testicular section of infected male rats treated with pyrimethamine (E) and sulphadiazine (F) showed intact seminiferous tubules and lumen full with sperms. The interstitial tissues also appeared intact with almost normal structure (40X, H and E stain).



A

B

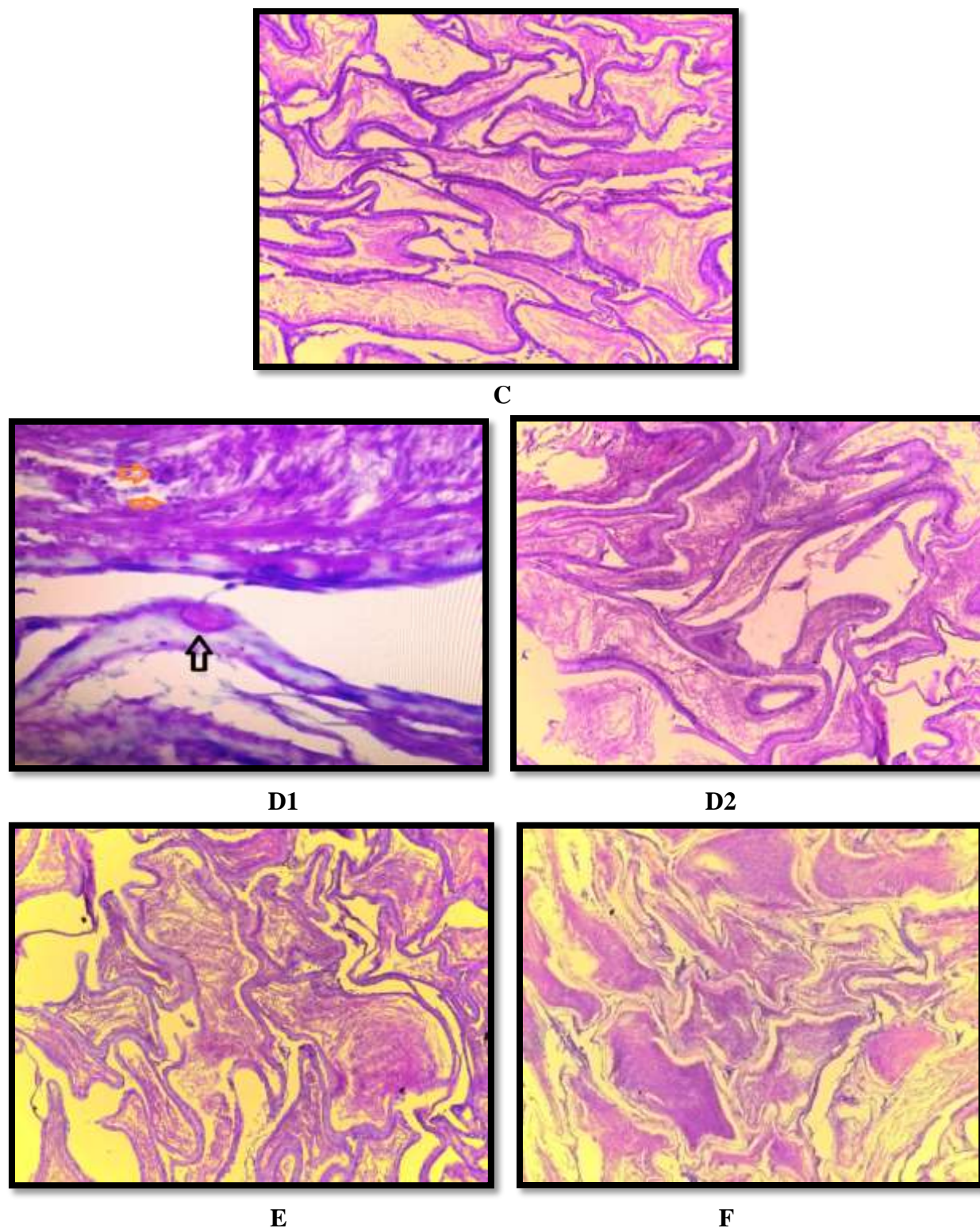


Figure 2: epididymal section of non infected male rats treated with DMSO

(A), pyrimethamine (B) and sulphadiazine (C) showed the normal histological structure. The epididymal section of infected male rats treated with DMSO showed hyperplasia of duct lining, sperms appeared impacted in the lumen (D1), with infiltration of inflammatory cells (→) and appearance of toxoplasma cysts (↑) (D2). The epididymal section of infected male rats treated

with pyrimethamine (E) and sulphadiazine (F) also showed hyperplasia of duct lining (40X, H and E stain).

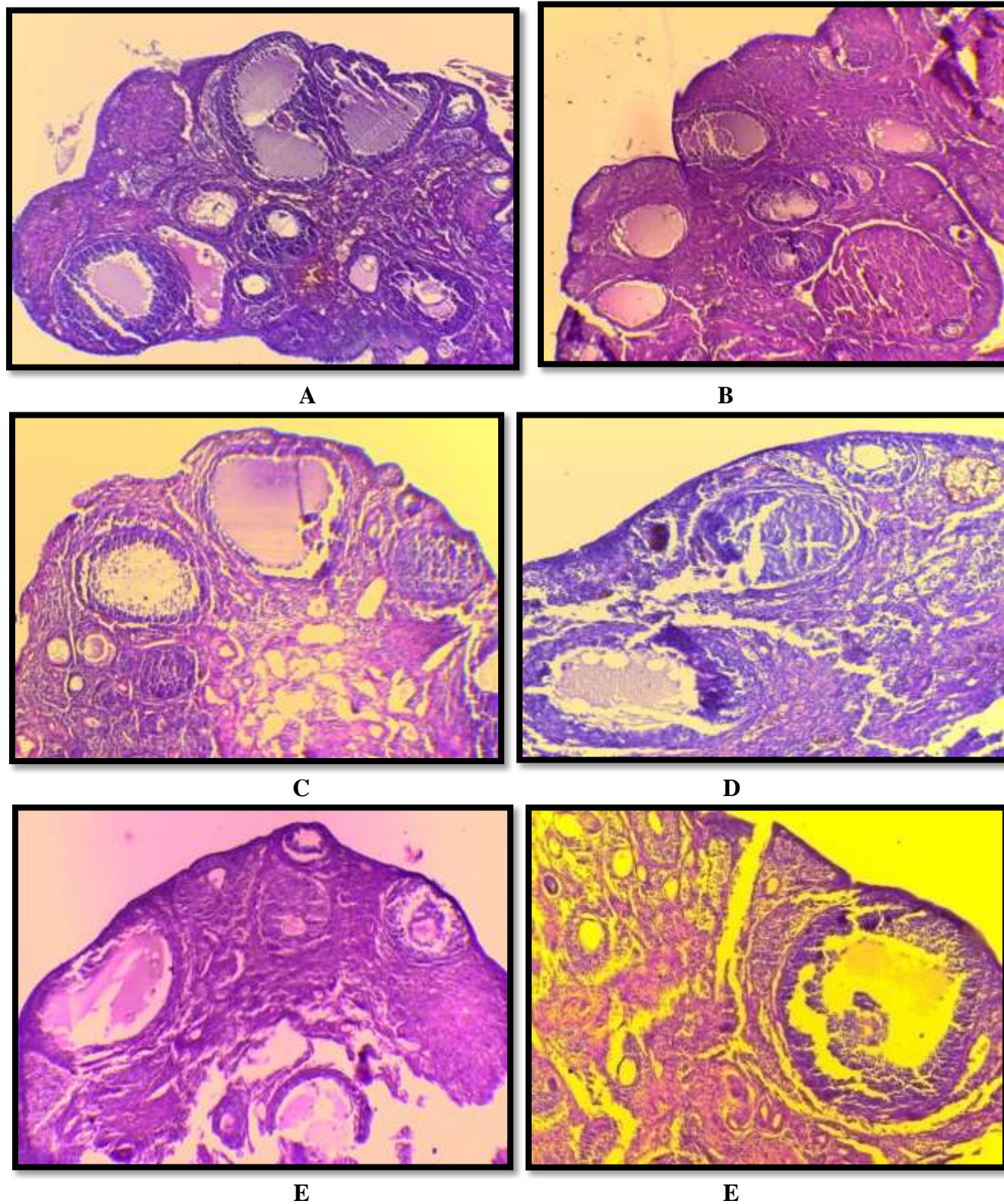
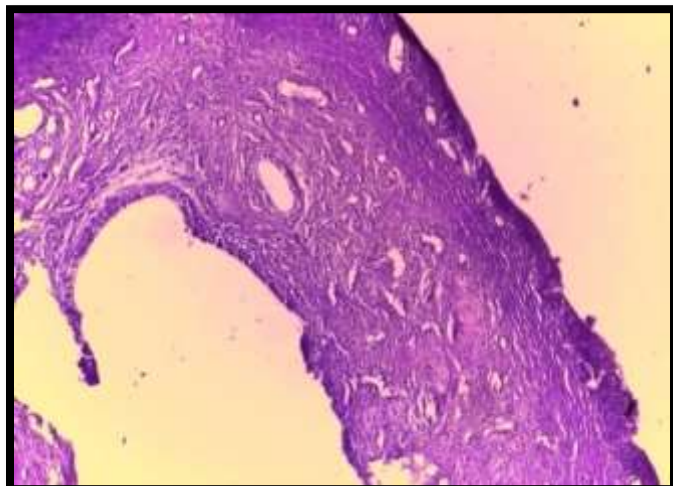
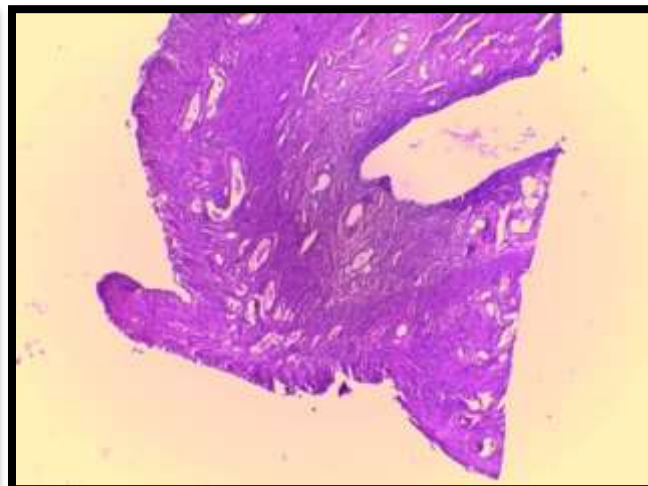
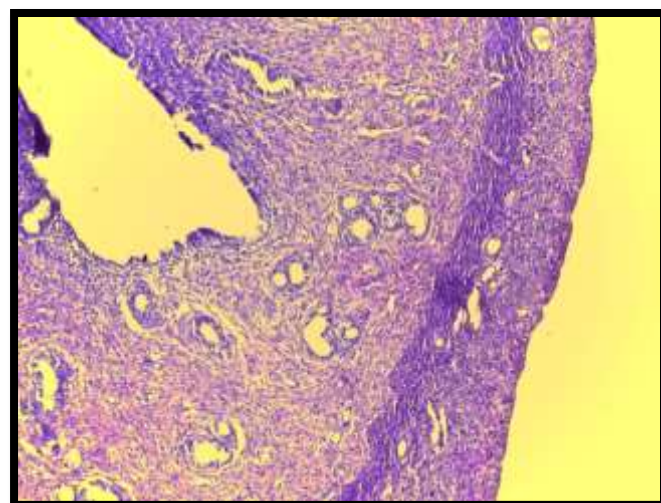
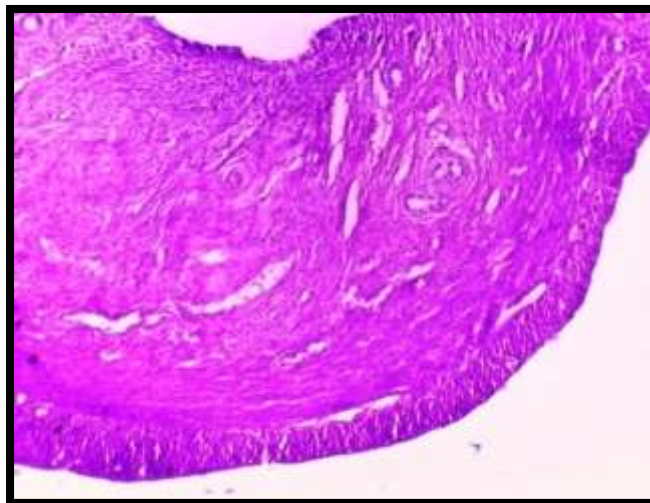
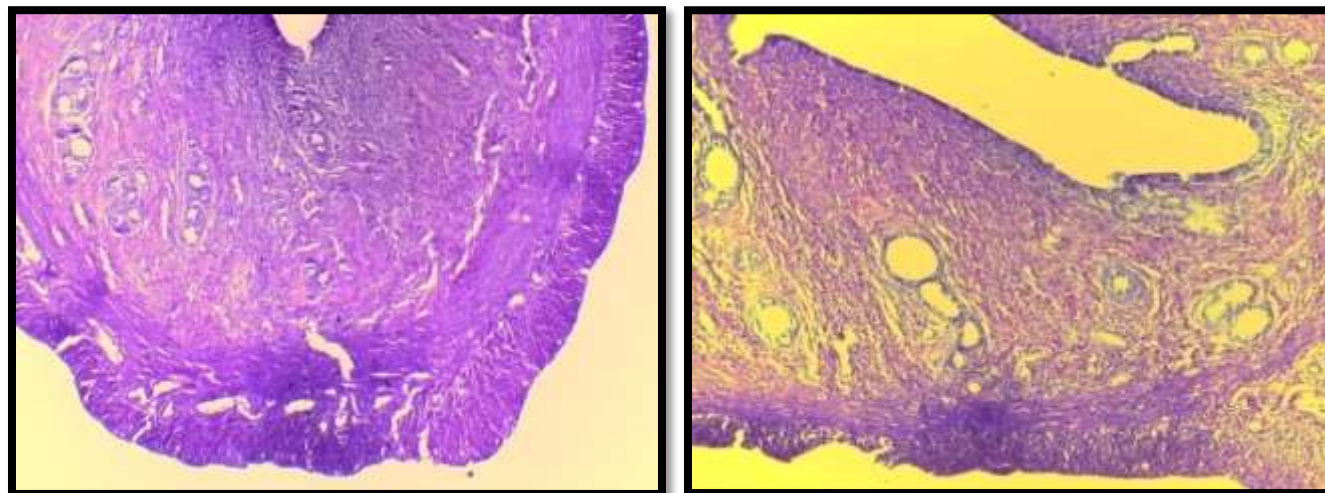


Figure 3: Ovarian section of non infected females treated by DMSO

(A), pyrimethamine (B) and sulphadiazine (C) showed the normal structure of the ovary. The histological section of the ovary of the infected female treated by DMSO showed decreased primary and secondary follicle with increased follicle atresia (D). The section of ovary of infected females treated by pyrimethamine (E) and sulphadiazine (E) showed that the structure and number of ovarian follicles return to normal limit (40X, H and E stain).

**A****B****C****D**



E

F

Figure 4: Uterine section of non infected females treated by DMSO

(A), pyrimethamine (B) and sulphadiazine (C) showed the normal structure of the uterus. The histological section of the uterus of the infected female treated by DMSO showed hypertrophy of the endometrium and myometrium, polymorphic inflammatory infiltration and fewer glands (D). The uterine sections of infected females treated by pyrimethamine (E) and sulphadiazine (E) also showed hypertrophy and increased thickness endometrium and myometrium (40X, H and E stain).

Our study revealed that the testis section of toxoplasma infected male rats showed vacular degeneration of spermatogonia and spermatocytes. The seminiferous tubules revealed severe degeneration, shrinkage, necrosis, hemorrhages with disappearance of epithelial lining of the majority of seminiferous tubules. They were also appeared separated with an irregular outlines and surrounded by fibrin. Seminiferous tubule lumen contained little amount of sperm with appearance of giant cells, polymorph nuclear leukocyte and exfoliated cellular debris. Sloughing of Leydig cells was also recorded. Furthermore, the epididymal sections of *Toxoplasma gondii* infected DMSO-treated male rats showed decreasing of the epithelial cell high and ducts diameters with infiltration of mononuclear and multinucleated giant cell. Epididymal sections also showed hyperplasia of duct lining, sperms appeared impacted in the lumen along with desquamated cellular debris. The histological changes found in testis and epididymis in this study were also recorded by Arantes *et al.*, the histopathological study of the reproductive system (testicles and epididymis) of male dogs experimentally infected with *Toxoplasma gondii* showed mononuclear inflammatory infiltrate in the epididymis, moderate cellular edema, hydropic degeneration and moderate interstitial fibrosis in seminiferous tubules¹⁶.

Lopes *et al.* investigated the reproductive histology in small male ruminants after *Toxoplasma gondii* infection. They noted diffuse testicular degeneration associated with calcification foci and a multifocal mononuclear interstitial inflammatory infiltrate; and a mononuclear interstitial infiltrate and focal necrotic areas of the muscle fibers surrounding the seminal vesicles¹⁷.

The histological changes in the ovary and uterus in *Toxoplasma gondii* infected female rats were also recorded previously. Eslamirad *et al.*, mentioned that the ovaries of *Toxoplasma gondii* infected pregnant mice showed gross morphological differences compared to the control groups. Primary follicles, secondary follicle and corpus luteum were significantly decreased in the infected females¹⁸. However, Fux *et al.*, recorded accentuated hypertrophy of the endometrium and myometrium and a reduction in folliculogenesis and formation of corpora lutea in the ovaries of *Toxoplasma gondii* infected mice¹⁹.

The histological changes could be attributed to the declined pituitary secretion and decreasing of its influence on male and female reproductive organs in the infected animals. Toxoplasmosis significantly decreased FSH and LH in both males and females, with a subsequent decline in the level of testosterone in males and estrogen and progesterone in females. Toxoplasmosis increased interleukin-1b (IL-1b) levels. The levels of IL-1b correlated significantly in a negative manner with FSH, LH, free testosterone (FT), total testosterone (TT) in all patients with acute toxoplasmosis¹⁵. Interleukin-1b suppressed the hypothalamic- pituitary- gonadal (HPG) axis, directly or indirectly through increased corticotrophin-releasing hormone (CRH) and/or cortisol²⁰. Interleukin-1b released peripherally in response to the parasite, reached the hypothalamus and initiated a sequence of events that inhibited the pulsatile release of gonadotropin-releasing hormone (GnRH), leading to the subsequent impairment of the pituitary-gonads axis⁶.

Our study also revealed that sulphadiazine and pyrimethamine didn't induce testicular, epididymal, ovarian and uterine histological changes in non infected males and females. The same results were recorded previously, Awoniyi *et al.*, mentioned that pyrimethamine 100 mg/kg/day did not alter the testicular and epididymal structure. Histological picture of the testis of the treated male was indistinguishable from controls²¹.

Many studies revealed that both drugs were free from endocrine effect, they didn't affected the hypothalamic – pituitary gonads axis both in males and females²¹⁻²², and didn't affected the relative weights of sexual and secondary sexual organ, semen quality and sexual performance, which could be explain the normal testicular, epididymal, ovarian and uterine histological structures of male and female rats treated by pyrimethamine and sulphadiazine in this study.

Furthermore, both sulphadiazine and pyrimethamine attenuated the histological changes associated with toxoplasma infection in both males and females. The attenuation of histological changes in infected male and female rats treated by sulphadiazine or pyrimethamine could be attributed to the therapeutic effects of these drugs in toxoplasmosis²³⁻²⁴.

CONCLUSION

Toxoplasmosis deteriorate the normal histological structure of sexual organs of both male and female rats. Pyrimethamine and sulphadiazine nor cause histological changed neither potentiate the histological effects of toxoplasmosis in infected and non infected male and female rats.

REFERENCES

1. Martinez-Garcia F, Regadera J, Mayer R, Sanchez S and Nistal M. Protozoan infections in the male genital tract. J Urol 1996; 156(2 Pt 1):340-349.
2. Nistal M, Santana A, Paniaqua R and Palacios J. Testicular toxoplasmosis in two men with the acquired immunodeficiency syndrome (AIDS). Arch Pathol Lab Med 1986; 110(8): 744-746.
3. De Paepe ME, Guerrieri C and Waxman M. Opportunistic infections of the testis in the acquired immunodeficiency syndrome. Mt Sinai J Med 1990; 57(1): 25-29.
4. Dvorakova-Hortova K, Sidlova A, Ded L, Hladovcova D, Vieweg M, Weidner W, Steger K, Stopka P and Dogan AP. *Toxoplasma gondii* decreases the reproductive fitness in mice. PLoS ONE 2004; 9(6): e96770.
5. Terpsidis KI, Papazahariadou M, Taitzoglou I, Papajoannou NG, Georgiadis MP and Theodoridis IT. *Toxoplasma gondii*: Reproductive parameters in experimentally infected male rats. Exp Parasitol 2009; 121(3): 238-241.
6. Stahl W, Dias J, Turek G and Kaneda G. Etiology of ovarian dysfunction in chronic murine toxoplasmosis. Parasitology Research 1995; 81(2):114-120.
7. Stahl W, Dias JA and Turek G. Hypothalamic-adenohypophyseal origin of reproductive failure in mice following chronic infection with *Toxoplasma gondii*. Proc Soc Exp Biol Med 1985; 178: 246-249.
8. Razzak AH, Wais SA and Saeid AY. Toxoplasmosis: the innocent suspect of pregnancy wastage in Duhok, Iraq. East Mediterr Health J 2005;11: 625-32.
9. Stahl W, Kaneda Y and Noguchi T. Reproductive failure in mice chronically infected with *Toxoplasma gondii*. Parasitol Res 1994; 80: 22-28.

10. Pal MN, Bhatia VN, Kotwani BG and Agarwal DS. Toxoplasmosis in relation to reproductive disorders. Ind J Med Res 1975; 63:11-16.
11. Oktenli C, Doganci L, Ozgurtas T, Araz RE, Tanyuksel M, Musabak U, Sanisoglu SY, Yesilova Z, Erbil MK and Inal A. Transient hypogonadotropic hypogonadism in males with acute toxoplasmosis: suppressive effect of interleukin-1b on the secretion of GnRH. Human Reprod 2004; 19: 859-866.
12. Zargar AH, Masoodi SR, Laway BA, Sofi BA and Wani AI. Seroprevalence of toxoplasmosis in women with repeated abortions in Kashmir. J Epidemiol Community Health 1998; 52: 135-136.
13. Al-Warid HS, Ali HZ and Muhamad SN. Detection of LTH, FSH and LH hormone level in pregnant women infected with *Toxoplasma gondii*. Int J of Recent Scientific Research 2012; 3(10): 809-811.
14. Lecomte V, Chumpitazi BFF, Pasquier B, Ambroise-Thomas P and Santoro F. Brain-tissue cysts in rats infected with the RH strain of *Toxoplasma gondii*. Parasitol Res 1992; 78: 267-269.
15. Inouye MJR. Differential staining of cartilage and bone in fetal mouse skeleton by alcian blue and alizarin reds. Cong Anom 1976;16: 171-173.
16. Arantes TP, Zanetti Lopes WDZ, Ferreira RM, Pieroni JP, Pinto VMR, Santos TR, Sakamoto CA and daCosta AJ. Histopathological analysis of the reproductive system of male dogs experimentally infected with *Toxoplasma gondii*. Ciência Rural 2009; 39(7): 2123-2127.
17. Lopes WD, Santos TR, Luvizotto MC, Sakamoto CA, Oliveira GP and Costa AJ. Histopathology of the reproductive system of male sheep experimentally infected with *Toxoplasma gondii*. Parasitol Res 2011;109(2):405-409.
18. Eslamirad Z, Bayat PD and Babaei S. Histological changes of the ovary in pregnant mice vaginally exposed to *Toxoplasma gondii*. Iranian Journal of Parasitology 2015; 10(2): 273-279.
19. Fux B, Ferreira AM, Cassali GD, Tafuri WL and Vitor RWA. Experimental toxoplasmosis in BALB/c mice. Prevention of vertical disease transmission by treatment and reproductive failure in chronic infection. Mem Inst Oswaldo Cruz, Rio de Janeiro 2000; 95(1): 121-126.
20. Rivier C, Rivier J and Vale W. Stress-induced inhibition of reproductive functions: role of endogenous corticotropin-releasing factor. Science 1986; 231: 607-609.

21. Awoniyi CA, Chandrashekar V, Hurst BS, Wookyoon K and Schlaff D. The effects of chronic administration of pyrimethamine on spermatogenesis and fertility in male rats. J Androl 1993; 14: 174-179.
22. Al- Snafi AE. Antimicrobial drugs. Al Dtaa Publication house, Iraq 2013.
23. Al- Snafi AE. Pharmacology and therapeutics. Al Dtaa Publication house, Iraq 2013.



AJPHR is
Peer-reviewed
monthly
Rapid publication
Submit your next manuscript at
editor@ajphr.com / editor.ajphr@gmail.com